



**UNIVERSITY OF MEDICINE AND PHARMACY  
"CAROL DAVILA", BUCHAREST**



**UNIVERSITY OF MEDICINE AND PHARMACY  
"CAROL DAVILA", BUCHAREST  
DOCTORAL SCHOOL  
PHARMACY FIELD**

**Implications of the properties of chitosan in the  
development of some intranasal insulin release systems  
targeting diseases at the CNS level**

**ABSTRACT OF THE DOCTORAL THESIS**

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## **Introduction**

The increase in the number of people diagnosed with diseases of the central nervous system motivated us to pay attention to the nasal administration of active pharmaceutical ingredients [1, 2]. This type of administration is less used, but has multiple advantages and needs to be explored in order to increase the bioavailability of active pharmaceutical ingredients. Up to date, a number of studies have been carried out for the intranasal administration of drugs with local and systemic action, but a greater interest is being allocated to those with action in the central nervous system.

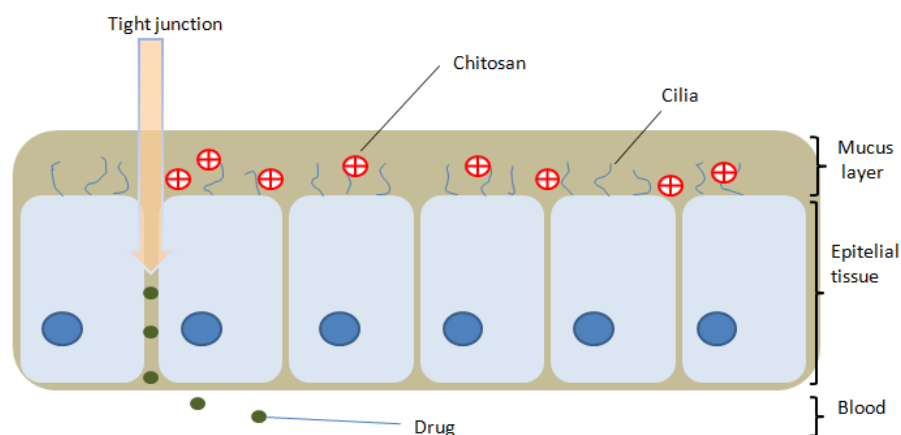
The well-known characteristics of chitosan and carboxymethyl chitosan polymers were the basis for the development of hydrocolloidal systems with nasal administration, because the impact of the mucoadhesive properties and of enhancing the permeability of the active pharmaceutical ingredient through the nasal mucosa is beneficial, thus favoring their bioavailability at the level of the central nervous system. The studies carried out so far have shown that the polymer and its derivatives are biodegradable, biocompatible and do not present toxicity, which makes them compatible with mucosal administration.

Insulin is a hormone, currently used to treat diabetes, but what is less known is that it also has actions at the level of the central nervous system. Insulin is considered a neuroprotein because it has been shown to have potentially beneficial therapeutic effects in conditions associated with the central nervous system, such as Alzheimer's disease, Parkinson's disease, and cognitive impairment. It also has the ability to modulate nicotine cravings and limit the symptoms associated with abstinence syndrome and favoring smoking cessation.

### **1. Chitosan. General aspects of the application of chitosan in the formulation of medicinal products**

Chitosan is a natural polysaccharide, generally obtained from a marine source [3]. Chitosan and its derivatives have mucoadhesive properties [4, 5], they enhance the permeability through the nasal mucosa due to the positive charge given by the amino groups, thus stimulating the bioavailability of the substances [6].

Regarding the surface tension of the polymer, it decreases with the increase in chitosan concentration [7, 8].



**Figure 1.1.** Mucoadhesive property of chitosan on the nasal mucosa and opening of the epithelial tight junction effect (Adapted after Popescu, R., M.V. Ghica, C.E. Dinu-Pirvu, V. Anuta, D. Lupuliasa, and L. Popa, New Opportunity to Formulate Intranasal Vaccines and Drug Delivery Systems Based on Chitosan. *Int J Mol Sci* 2020, 21 [6])

The interactions between chitosan and proteins [9], lipids [10] or other molecules [11] lead to compounds that have their own applications or to the modification of the characteristics of the substances involved in the connection (conformational changes [12], increased solubility [13], etc.).

Chitosan and its derivatives have their own actions (antibacterial [14, 15], antifungal [16], antitumor [17], antioxidant [18], hemostatic [19], immunostimulatory [20], neuroprotective [21] and others) and can be used as such or in combination with other molecules of the same class to achieve a synergistic effect [22-24].

Studying the properties of chitosan and its derivatives is necessary to evaluate their influence in the development of nasal formulations. Studies to date have shown that chitosan is biodegradable, biocompatible and non-toxic making it suitable for mucosal delivery, leading to the formulation of nasal drug delivery systems [25-27].

## 2. Chitosan-Based Formulations for Intranasal Administration

The nasal route shows higher bioavailability compared to the oral route, avoiding hepatic [28] and intestinal metabolism [29]. Biologically active drugs reach the brain directly, from the nasal cavity, along the trigeminal nerve and through the olfactory system, bypassing the blood-brain barrier in a non-invasive way [28, 30], or are absorbed through the nasal epithelium into the systemic circulation and then they pass through the BBB into the CNS [31]. Systems that transport the biologically active molecules in the body without being invasive and without penetrating the BBB are considered third generation [32].

Nasal vaccine can be used to overcome the disadvantages of the conventional and injectable ones [30]. These can prevent outbreaks or pandemics, leading to a more effective management of transmissible diseases. The use of nasal vaccines can increase the percentage of people immunized [33]. As the nasal route is non-invasive and rapid [34], the vaccine can be easily administered at home without the need for qualified medical personnel [28]. The use of chitosan as an adjuvant in the formulation of nasal vaccines helps to obtain a superior immune response [35].

According to the literature, numerous researches include drug delivery systems containing chitosan or its derivatives for intranasal administration and incorporating different active substances, such as: hydrogels with ropinirole or insulin [36], nanoparticles with methylprednisone or insulin [37], microspheres with carvedilol or diltiazem [38], emulsions with quetiapine or zolmitriptan [39] and so on.

Besides the well-known property of insulin to regulate glucose metabolism in the treatment of diabetes [39, 40], intranasal administration also shows favorable effects in CNS-related diseases [41, 42] through its neuroprotective action [43]. Different clinical studies aim to investigate the potential therapeutic effects of insulin in diseases such as: memory impairment, Alzheimer's disease, Parkinson's disease, smoking cessation and others [44].

### **3. Working hypothesis and general objectives**

The research presented in the present thesis was based on the biodegradable, biocompatible and non-toxic characteristics of chitosan, but most importantly, on the mucoadhesive properties and its ability to improve the penetration through the nasal mucosa of active pharmaceutical ingredient in general, but especially of insulin.

Studies carried out to date have shown the benefits of intranasal insulin administration, and in this work, we aim to evaluate the properties of hydrocolloidal systems based on chitosan and its derivatives containing insulin.

The main objective of the doctoral work is represented by the formulation and evaluation of hydrocolloidal systems based on chitosan and, respectively, using the derivative - carboxymethyl chitosan, which incorporates insulin, with the aim of transporting the active substance from the nose to the brain.

### **4. General research methodology**

The general research methodology, which was developed within the collective of the Physico-Chemical and Colloidal Discipline of the Faculty of Pharmacy, within the "Carol Davila" University of Medicine and Pharmacy in Bucharest.



The pH was evaluated using the METTLER TOLEDO Seven Compact pH-meter. The calibration of the device was carried out with a standard solution with pH 4, then with pH 7 before making the measurements.

The evaluation of the surface properties consisted in the determination of the contact angle by the sessile drop method and the surface tension by the falling drop method using the goniometer CAM 101 (KSV Instruments, Finland) based on the Young-Laplace equation. The results obtained from the two determinations were included in the calculation of the work of adhesion, the work of cohesion and the spreading coefficient of the hydrocolloidal systems.

Determination of the rheological profile of the systems was performed with the Multi Visc viscometer (Fungilab SA, Barcelona, Spain), using the adapter for reduced viscosities for chitosan and PVA-based systems. The experiments were carried out at the conditioning temperature (4 – 8 °C), and the mathematical modeling of the rheological profiles was used to determine the viscosity. For the second study, the Lamy Rheology RM100 Plus viscometer (Lamy Rheology Instruments, Champagne au Mont d'Or, France) was used, at a temperature of 35 °C. The Power Law Model was applied to analyze the flow behavior.

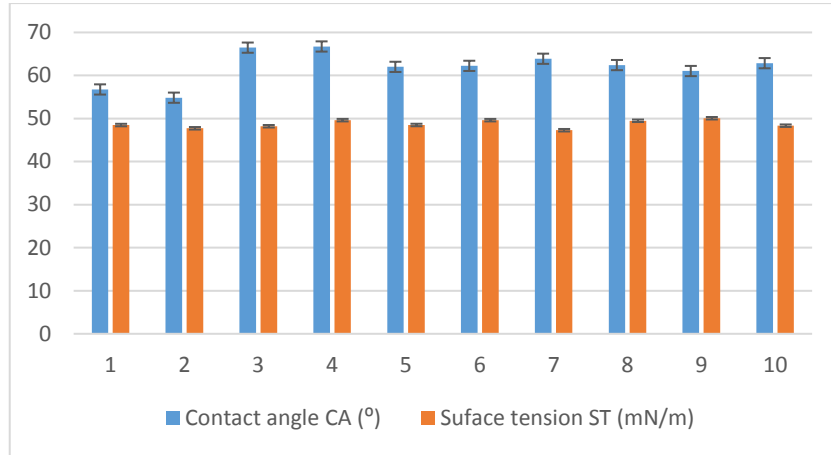
The *in vitro* release studies, corresponding to the second experiment, were performed on Franz diffusion cells (Hanson apparatus), using synthetic cellulose acetate membrane, using phosphate buffer, at 35°C. At predefined time intervals, samples were taken, which were analyzed spectrophotometric at 271 nm.

## **5. Development and preliminary evaluation of chitosan-based intranasal hydrocolloidal systems with insulin**

The first study involved the development, formulation and preliminary evaluation of intranasal hydrocolloid systems based on chitosan and polyvinyl alcohol with insulin. Research was focused on determining the surface properties of the hydrocolloid systems, especially mucoadhesiveness; as well as rheological profiles, as key factors to overcome mucociliary clearance and improve therapeutic efficacy after intranasal drug administration.

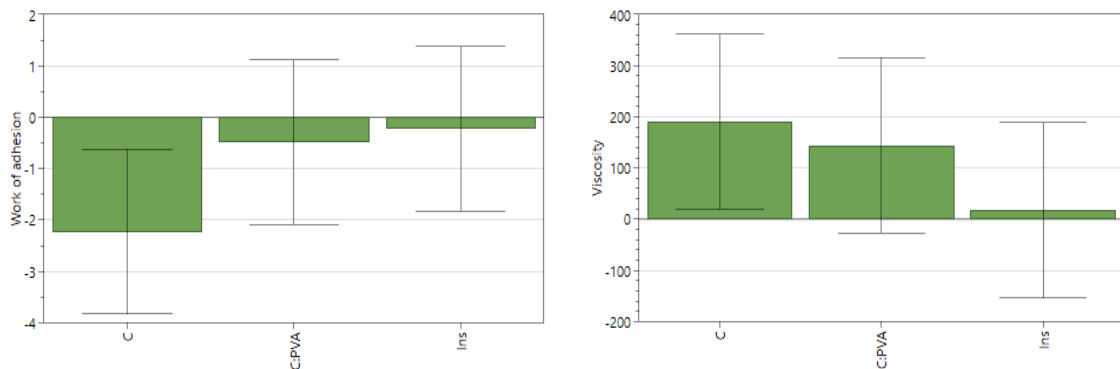
To design the experimental matrix in this study, the Modde 13 program was used, in which there are 3 independent variables X1: chitosan concentration (1%, 1.5%, 2%); X2: ratio between chitosan and PVA 1% (1:2, 1:1, 2:1) and X3: amount of insulin (5 IU/g, 7.5 IU/g, 10 IU/g) [45]. In addition, the Minitab statistical program was used to generate the regression equations for each response.

The contact angle values for the hydrocolloidal systems analyzed in this study were between  $54.84 \pm 1.52^\circ$  and  $66.74 \pm 0.59^\circ$ ; which indicates an increased wetting capacity and a good spreading capacity of the systems. The surface tension values obtained from the determinations are lower than that of the nasal mucosa [4].



**Figure 5.1.** Comparative graph of contact angle and surface tension

According to the graphical representation of the regression coefficients, the increase in chitosan concentration has an inversely proportional influence on the adhesion forces and proportional to the viscosity.



**Figure 5.2.** Regression coefficient plots for to the work of adhesion and viscosity

Putting together the results of the study, regarding the superficial properties and rheological profiles, the hydrocolloidal system 1 (chitosan 1%, chitosan:PVA = 2:1, insulin = 5 IU/g) can be considered the optimal sample and can be used for further research.

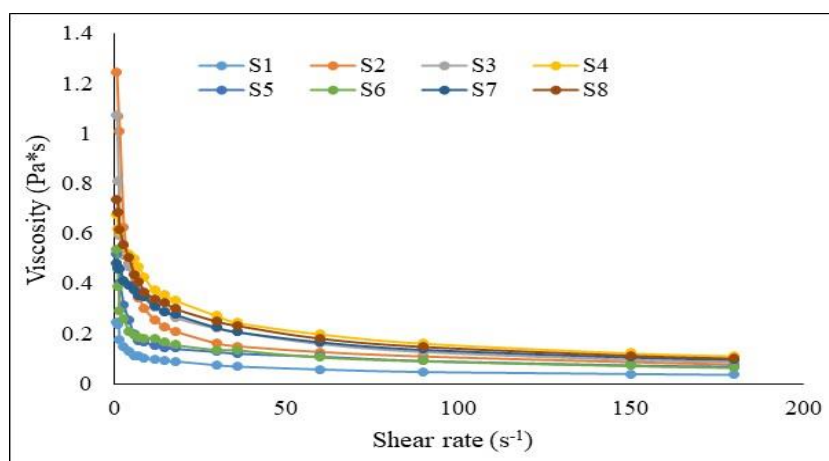
## 6. Physico-chemical characterization and initial evaluation of intranasal hydrocolloid insulin systems based on carboxymethyl chitosan – hyaluronan

The aim of this study was to develop and characterize nasal systems based on carboxymethyl chitosan (the soluble derivative of chitosan [46]) and sodium hyaluronate

with insulin, which would combine the mucoadhesive properties of the polymers and stimulate insulin absorption [47-49]. We evaluated the influence of formulation factors on the superficial properties, rheology and *in vitro* release of insulin using complementary Design of Experiments strategies.

A factorial design of  $2^3$  was developed, where there are three independent variables at two levels of variation, where X1 is the concentration of CMC (1% or 2%), X2 is the ratio of CMC to NaHA (1/1 or 1/2) and X3 is the amount of insulin (20 IU/ml or 30 IU/ml).

All studied samples recorded the contact angle values lower than  $90^\circ$ . The determinations made in this experiment showed that formulation S1 had the closest value to the physiological surface tension of the nasal mucosa. The work of adhesion calculated based on contact angle and surface tension was over 80 mN/m for all systems. The flow behavior of the eight systems was pseudoplastic.

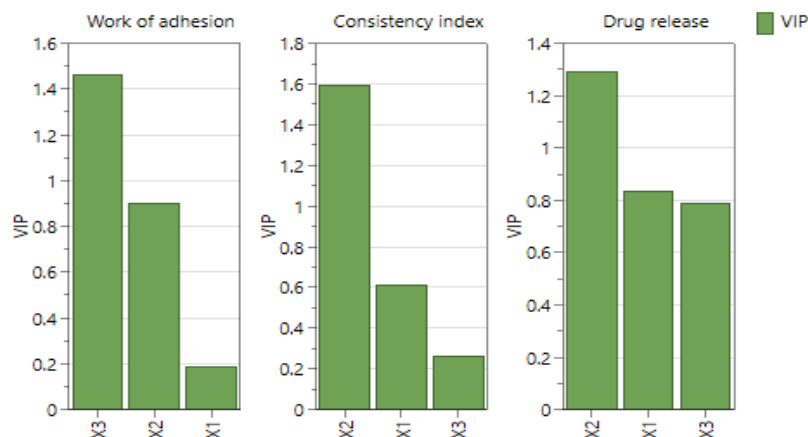


**Figure 6.1.** The influence of share rate on the viscosity

The release kinetics followed the zero-order model for S1, S4, S7 and S8 with  $R^2$  greater than 0,9934 and the rest of the systems the Power-law model with  $R^2 > 0,9958$ ; and all samples had cumulative drug release higher than 60%.

Modde software was used to design the experimental plan and provide 3D representation of the response surface and response contour, to observe the trend of the response variables based on the changes in the formulation parameter [50]. Minitab statistical software was used to analyze the data and complement the screening provided by the Modde program by generating regression equations and Pareto charts for each response factor.

The work of adhesion was influenced by the amount of insulin (X3); and by the CMC/NaHA ratio (X2). The CMC/NaHA ratio had pronounced significance in terms of the consistency index and the release of the amount of active substance.



**Figure 6.2.** Variable importance in the projection (VIP) of the independent parameters

The S7 system is the most eloquent system compared to the predictive point of reference generated with the Modde program.

## 7. Conclusions and personal contributions

The doctoral thesis, entitled "*Implications of the properties of chitosan in the development of some intranasal insulin release systems targeting diseases at the CNS level*" aims to overcome the limitations given by the blood-brain barrier on active pharmaceutical ingredients acting on the central nervous system, by outlining the characteristics of chitosan and its derivatives in the formulation of medicinal systems for nasal administration.

The objectives proposed in this doctoral thesis were met, and that are: the selection of a medicinal substance with new applications discovered within the diseases associated with the central nervous system; the formulation and development of two hydrocolloid systems based on chitosan or carboxymethyl chitosan associated with other polymers, to enhance the properties of the polymers; evaluating the impact of the formulation parameters on the response variables with the help of two statistical programs. These interpretations can be used as a starting point for further studies.

These hydrocolloid systems can be included in other researches, for administration on the nasal mucosa or with applicability on other mucous membranes and in which to incorporate other active substances from different drug classes, offering new opportunities for treatment.

## Bibliography

1. Mureșanu DF, Chira D, Dobran Ș-A, Gherman A. The 2nd edition of the Romanian National Neurology Forum: from idea to implementation in the health system – here, now, together! *Journal of Medicine and Life*. 2024;17(2):129-32.
2. Collaborators GBDNSD. Global, regional, and national burden of disorders affecting the nervous system, 1990-2021: a systematic analysis for the Global Burden of Disease Study 2021. *Lancet Neurol*. 2024;23(4):344-81.
3. TM MW, Lau WM, Khutoryanskiy VV. Chitosan and Its Derivatives for Application in Mucoadhesive Drug Delivery Systems. *Polymers (Basel)*. 2018;10(3):1-37.
4. Mati-Baouche N, Elchinger P-H, de Baynast H, Pierre G, Delattre C, Michaud P. Chitosan as an adhesive. *European Polymer Journal*. 2014;60:198-212.
5. Popa L, Ghica MV, **Popescu R**, Irimia T, Dinu-Pîrvu C-E. Development and Optimization of Chitosan-Hydroxypropyl Methylcellulose In Situ Gelling Systems for Ophthalmic Delivery of Bupivacaine Hydrochloride. *Processes*. 2021;9(10).
6. **Popescu R**, Ghica MV, Dinu-Pîrvu CE, Anuta V, Lupuliasa D, Popa L. New Opportunity to Formulate Intranasal Vaccines and Drug Delivery Systems Based on Chitosan. *Int J Mol Sci*. 2020;21(14).
7. Elsabee MZ, Morsi RE, Al-Sabagh AM. Surface active properties of chitosan and its derivatives. *Colloids Surf B Biointerfaces*. 2009;74(1):1-16.
8. Nilsen-Nygaard J, Strand S, Vårum K, Draget K, Nordgård C. Chitosan: Gels and Interfacial Properties. *Polymers*. 2015;7(3):552-79.
9. Azevedo JR, Sizilio RH, Brito MB, Costa AMB, Serafini MR, Araújo AAS, et al. Physical and chemical characterization insulin-loaded chitosan-TPP nanoparticles. *Journal of Thermal Analysis and Calorimetry*. 2011;106(3):685-9.
10. Vemireddy S, M CP, Halmuthur MS. Chitosan stabilized nasal emulsion delivery system for effective humoral and cellular response against recombinant tetravalent dengue antigen. *Carbohydr Polym*. 2018;190:129-38.
11. Fini A, Orienti I. The Role of Chitosan in Drug Delivery. *American Journal of Drug Delivery*. 2003;1(1):43-59.
12. Shariatinia Z. Pharmaceutical applications of chitosan. *Adv Colloid Interface Sci*. 2019;263:131-94.
13. Boonsongrit Y, Mitrevej A, Mueller BW. Chitosan drug binding by ionic interaction. *Eur J Pharm Biopharm*. 2006;62(3):267-74.

14. Benltoufa S, Miled W, Trad M, Slama RB, Fayala F. Chitosan hydrogel-coated cellulosic fabric for medical end-use: Antibacterial properties, basic mechanical and comfort properties. *Carbohydr Polym.* 2020;227:115352.
15. Islam MM, Islam R, Mahmudul Hassan SM, Karim MR, Rahman MM, Rahman S, et al. Carboxymethyl chitin and chitosan derivatives: Synthesis, characterization and antibacterial activity. *Carbohydrate Polymer Technologies and Applications.* 2023;5.
16. Munoz-Tebar N, Perez-Alvarez JA, Fernandez-Lopez J, Viuda-Martos M. Chitosan Edible Films and Coatings with Added Bioactive Compounds: Antibacterial and Antioxidant Properties and Their Application to Food Products: A Review. *Polymers (Basel).* 2023;15(2).
17. Ju H, Yu C, Liu W, Li H-H, Fu Z, Wu Y-C, et al. Polysaccharides from marine resources exhibit great potential in the treatment of tumor: A review. *Carbohydrate Polymer Technologies and Applications.* 2023;5.
18. Desai N, Rana D, Salave S, Gupta R, Patel P, Karunakaran B, et al. Chitosan: A Potential Biopolymer in Drug Delivery and Biomedical Applications. *Pharmaceutics.* 2023;15(4).
19. Zhang Hu, Sitong Lu, Yu Cheng, Songzhi Kong, Sidong Li, Chengpeng Li, et al. Investigation of the Effects of Molecular Parameters on the Hemostatic Properties of Chitosan. *Molecules.* 2018;23:1-14.
20. Li M, Wang Y, Sun Y, Cui H, Zhu SJ, Qiu HJ. Mucosal vaccines: Strategies and challenges. *Immunol Lett.* 2020;217:116-25.
21. Ojeda-Hernandez DD, Canales-Aguirre AA, Matias-Guiu J, Gomez-Pinedo U, Mateos-Diaz JC. Potential of Chitosan and Its Derivatives for Biomedical Applications in the Central Nervous System. *Front Bioeng Biotechnol.* 2020;8:389.
22. Yu Z, Rao G, Wei Y, Yu J, Wu S, Fang Y. Preparation, characterization, and antibacterial properties of biofilms comprising chitosan and epsilon-polylysine. *Int J Biol Macromol.* 2019;141:545-52.
23. Julia Radwan-Pragłowska, Marek Pi, Volodymyr Deineka, Łukasz Janus, Viktoriia Korniienko, Evgenia Husak, et al. Chitosan-Based Bioactive Hemostatic Agents with Antibacterial Properties—Synthesis and Characterization. *Molecules.* 2019;24:1-17.
24. Yingying Hua, Chenjun Ma, Tiantian Wei, Zhang L, Shen J. Collagen/Chitosan Complexes: Preparation, Antioxidant Activity, Tyrosinase Inhibition Activity, and Melanin Synthesis. *International Journal of Molecular Sciences.* 2020;21:1-15.

25. Martau GA, Mihai M, Vodnar DC. The Use of Chitosan, Alginate, and Pectin in the Biomedical and Food Sector-Biocompatibility, Bioadhesiveness, and Biodegradability. *Polymers (Basel)*. 2019;11(11):1-28.
26. George A, Shah PA, Shrivastav PS. Natural biodegradable polymers based nano-formulations for drug delivery: A review. *Int J Pharm*. 2019;561:244-64.
27. Upadhyaya L, Singh J, Agarwal V, Tewari RP. Biomedical applications of carboxymethyl chitosans. *Carbohydr Polym*. 2013;91(1):452-66.
28. Pacheco C, Sousa F, Sarmiento B. Chitosan-based nanomedicine for brain delivery: Where are we heading? *Reactive and Functional Polymers*. 2020;146:104430.
29. Fan LW, Carter K, Bhatt A, Pang Y. Rapid transport of insulin to the brain following intranasal administration in rats. *Neural Regen Res*. 2019;14(6):1046-51.
30. Wang Q, Zuo Z, Cheung CKC, Leung SSY. Updates on thermosensitive hydrogel for nasal, ocular and cutaneous delivery. *Int J Pharm*. 2019;559:86-101.
31. Khosa A, Saha RN, Singhvi G. Drug delivery to the brain. *Nanomaterials for Drug Delivery and Therapy*2019. p. 461-514.
32. Pandey SP, Shukla T, Dhote VK, K. Mishra D, Maheshwari R, Tekade RK. Use of Polymers in Controlled Release of Active Agents. *Basic Fundamentals of Drug Delivery*2019. p. 113-72.
33. Bhavana V, Thakor P, Singh SB, Mehra NK. COVID-19: Pathophysiology, treatment options, nanotechnology approaches, and research agenda to combating the SARS-CoV2 pandemic. *Life Sci*. 2020;261:118336.
34. Lochhead JJ, Kellohen KL, Ronaldson PT, Davis TP. Distribution of insulin in trigeminal nerve and brain after intranasal administration. *Sci Rep*. 2019;9(1):2621.
35. Wu D, Zhu L, Li Y, Zhang X, Xu S, Yang G, et al. Chitosan-based Colloidal Polyelectrolyte Complexes for Drug Delivery: A Review. *Carbohydrate Polymers*. 2020:116126.
36. Liu L, Gao Q, Lu X, Zhou H. In situ forming hydrogels based on chitosan for drug delivery and tissue regeneration. *Asian Journal of Pharmaceutical Sciences*. 2016;11(6):673-83.
37. Ansari R, Sadati SM, Mozafari N, Ashrafi H, Azadi A. Carbohydrate polymer-based nanoparticle application in drug delivery for CNS-related disorders. *European Polymer Journal*. 2020;128:109607.

38. Tiozzo Fasiolo L, Manniello MD, Tratta E, Buttini F, Rossi A, Sonvico F, et al. Opportunity and challenges of nasal powders: Drug formulation and delivery. *Eur J Pharm Sci.* 2018;113:2-17.
39. Awad R, Avital A, Sosnik A. Polymeric nanocarriers for nose-to-brain drug delivery in neurodegenerative diseases and neurodevelopmental disorders. *Acta Pharmaceutica Sinica B.* 2022.
40. \*\*\*. The United States Pharmacopeia and National Formulary. USP 36, NF 31. 2013.
41. Santiago JCP, Hallschmid M. Outcomes and clinical implications of intranasal insulin administration to the central nervous system. *Exp Neurol.* 2019;317:180-90.
42. Wingrove J, Swedrowska M, Scherliess R, Parry M, Ramjeeawon M, Taylor D, et al. Characterisation of nasal devices for delivery of insulin to the brain and evaluation in humans using functional magnetic resonance imaging. *J Control Release.* 2019;302:140-7.
43. Talati CP, Lee JW, Lu S, Ojeda NB, Prakash V, Dankhara N, et al. Intranasal insulin attenuates hypoxia-ischemia-induced short-term sensorimotor behavioral disturbances, neuronal apoptosis, and brain damage in neonatal rats. *Curr Res Neurobiol.* 2024;6:100123.
44. ClinicalTrials.gov. <https://www.clinicaltrials.gov>. accessed on 10.07.2024.
45. **R. Popescu**, C-E Dinu-Pîrvu, M.V. Ghica, V. Anuța, L. Popa. Development and preliminary evaluation of intranasal hydrocolloidal systems based on chitosan and PVA with insulin, for central nervous system-associated diseases. *Farmacia.* 2024;72:963-74.
46. Pang HT, Chen XG, Park HJ, Cha DS, Kennedy JF. Preparation and rheological properties of deoxycholate-chitosan and carboxymethyl-chitosan in aqueous systems. *Carbohydrate Polymers.* 2007;69(3):419-25.
47. Wong CYJ, Baldelli A, Hoyos CM, Tietz O, Ong HX, Traini D. Insulin Delivery to the Brain via the Nasal Route: Unraveling the Potential for Alzheimer's Disease Therapy. *Drug Deliv Transl Res.* 2024.
48. Mourya VK, Inamdara N, Ashutosh Tiwari N. Carboxymethyl Chitosan And Its Applications. *Advanced Materials Letters.* 2010;1(1):11-33.
49. Cuomo F, de Nigris A, Zeppa L, Lopez F, Ambrosone L. Viscosimetric properties of sodium hyaluronate and hypromellose solutions for medical devices. *Journal of Molecular Liquids.* 2024;398.
50. **Popescu R**, Dinu-Pîrvu C-E, Ghica MV, Anuța V, Popa L. Physico-Chemical Characterization and Initial Evaluation of Carboxymethyl Chitosan–Hyaluronan Hydrocolloid Systems with Insulin Intended for Intranasal Administration. *International Journal of Molecular Sciences.* 2024;25(19):10452.



## List of scientific papers developed in relation to the research topic

### ARTICLES PUBLISHED IN ISI INDEXED JOURNALS WITH IMPACT FACTOR

1. **Popescu R**, Dinu-Pîrvu C-E, Ghica MV, Anuța V, Popa L. Physico-Chemical Characterization and Initial Evaluation of Carboxymethyl Chitosan–Hyaluronan Hydrocolloid Systems with Insulin Intended for Intranasal Administration. *International Journal of Molecular Sciences*, 2024, 25(19), 10452.

<https://doi.org/10.3390/ijms251910452> , ISSN 1422-0067, IF – 4,9/2023.

<https://www.mdpi.com/1422-0067/25/19/10452> (Chapter 6).

2. **R. Popescu**, C.E. Dinu-Pîrvu, M.V. Ghica, V. Anuța, L. Popa, *Development and preliminary evaluation of intranasal hydrocolloidal systems based on chitosan and PVA with insulin, for central nervous system-associated diseases*. *Farmacia* **2024**, 72(4), 963-974.

<https://doi.org/10.31925/farmacia.2024.4.25>, ISSN 2065-0019, IF - 1,6/2022

<https://farmaciajournal.com/issue-articles/development-and-preliminary-evaluation-of-intranasal-hydrocolloidal-systems-based-on-chitosan-and-pva-with-insulin-for-central-nervous-system-associated-diseases/> (Chapter 5).

3. Popa, L., M.V. Ghica, **R. Popescu**, T. Irimia, and C.-E. Dinu-Pîrvu, *Development and Optimization of Chitosan-Hydroxypropyl Methylcellulose In Situ Gelling Systems for Ophthalmic Delivery of Bupivacaine Hydrochloride*. *Processes* **2021**, 9(10), 1694.

<https://doi.org/10.3390/pr9101694>, ISSN 2227-9717, IF – 2,8/2023.

<https://www.mdpi.com/2227-9717/9/10/1694>

4. **Popescu, R.**, M.V. Ghica, C.E. Dinu-Pîrvu, V. Anuta, D. Lupuliasa, and L. Popa, *New Opportunity to Formulate Intranasal Vaccines and Drug Delivery Systems Based on Chitosan*. *Int J Mol Sci* **2020**, 21(14), 5016.

<https://doi.org/10.3390/ijms21145016>, ISSN 1422-0067, IF - 4.9/2023

<https://www.mdpi.com/1422-0067/21/14/5016> (Chapter 1 and 2)

**STUDIES PUBLISHED IN SUMMARY IN JOURNALS AND IN THE  
VOLUMES OF SOME INTERNATIONAL SCIENTIFIC EXHIBITIONS**

1. Popa L., **R. Popescu**, M.V. Ghica, C.E. Dinu-Pîrvu, V. Anuța, M.T. Talianu, *Unveiling chitosan-based hydrocolloidal systems incorporating insulin for nasal administration*, poster paper (Poster 7, Board A - Section Formulation 2) presented at Congressus Pharmaceuticus Hungaricus (CPH) XVII and EUFEPS Annual Meeting, Debrecen, Hungary, 23<sup>th</sup> – 25<sup>th</sup> May 2024, volume of summaries, 2024, p. 384-385.

[https://clubservice-event.hu/pdf-egyeb/abstracts\\_CPH2024.pdf](https://clubservice-event.hu/pdf-egyeb/abstracts_CPH2024.pdf)

**STUDIES PUBLISHED IN SUMMARY IN JOURNALS AND IN THE  
VOLUMES OF SOME NATIONAL SCIENTIFIC EXHIBITIONS WITH  
ISBN/ISSN**

1. **R. Popescu**, L. Popa, M.V. Ghica, C.E. Dinu-Pîrvu, V. Anuța, R.M. Prisada, *Applying chitosan properties in designing of innovative systems for intranasal insulin delivery*, e-poster paper (ID 711), presented at "Carol Davila" University of Medicine and Pharmacy Congress, 11<sup>th</sup> edition, Bucharest, October 26-28, 2023, volume of abstracts: Supplement Maedica - a Journal of Clinical Medicine, 2023, Vol.18, p. 14, ISSN 2501-6903.

<https://www.congresumf.ro/supliment-maedica/>

<https://www.congresumf.ro/editia-2023/>

2. **R. Popescu**, L. Popa, M.V. Ghica, C.E. Dinu-Pîrvu, V. Anuța, R.M. Prisada, *Chitosan biopolymer in pharmaceutical systems with mucosal administration: evolution, challenges and perspectives*, oral communication paper presented during the 100<sup>th</sup> anniversary of the establishment of the autonomous Pharmaceutical University Education in Romania - Celebration of the Centenary of the Faculty of Pharmacy in Bucharest, June 12-15, 2023, Bucharest, volume of abstracts, p. 90, ISSN 2457-3027.